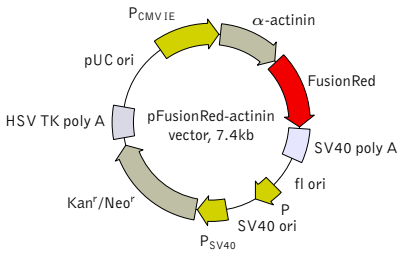
pFusionRed-actinin vector

The vector sequence has been compiled using the information from sequence databases, published literature, and other sources, together with partial sequences obtained by Evrogen. This vector has not been completely sequenced.



For vector sequence, please visit our Web site at http://www.evrogen.com/products/vectors.shtml

Product	Cat.#	Size
pFusionRed-actinin vector	**FP426**	20 μg

Vector type	mammalian expression vector
Reporter	FusionRed
Reporter codon usage	mammalian
Promoter for FusionRed	$P_{CMV\ IE}$
Host cells	mammalian
Selection	prokaryotic - kanamycin eukaryotic - neomycin (G418)
Replication	prokaryotic - pUC ori eukaryotic - SV40 ori
Use	red fluorescent labeling of α-actinin

Location of features

$P_{CMV\ IE}$: 1-589
Enhancer region: 59-465
TATA box: 554-560
Transcription start point: 583
alpha-actinin-FusionRed fusion: 637-4068
alpha-actinin: 637-3312
Start codon (ATG): 637-639
Last amino acid in alpha-actinin: 3310-3312
FusionRed: 3370-4068
Stop codon: 4066-4068
SV40 early mRNA polyadenylation signal
Polyadenylation signals: 4221-4226 & 4250-4255
mRNA 3' ends: 4259 & 4271
f1 single-strand DNA origin: 4318-4773
Bacterial promoter for expression of Kan^r gene
-35 region: 4835-4840; -10 region: 4858-4863
Transcription start point: 4870
SV40 origin of replication: 5114-5249
SV40 early promoter
Enhancer (72-bp tandem repeats): 4947-5018 & 5019-5090
21-bp repeats: 5094-5114, 5115-5135 & 5137-5157
Early promoter element: 5170-5176
Major transcription start points: 5166, 5204, 5210 & 5215
Kanamycin/neomycin resistance gene
Neomycin phosphotransferase coding sequences:
Start codon (ATG): 5298-5300; Stop codon: 6090-6092
G->A mutation to remove Pst I site: 5480
C->A (Arg to Ser) mutation to remove BssH II site: 5826
Herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signal
Polyadenylation signals: 6328-6333 & 6341-6346
pUC plasmid replication origin: 6677-7320

Vector description

pFusionRed-actinin is a mammalian expression vector encoding FusionRed-actinin fusion protein. The vector can be used for fluorescent labeling of α-actinin in living cells.

FusionRed codon usage is optimized for high expression in mammalian cells (humanized) [Haas et al. 1996]. Human α-actinin is fused to the FusionRed N-terminus.

pFusionRed-actinin vector can be used as a source of FusionRed-actinin hybrid sequence. The vector backbone contains unique restriction sites that permit its excision and further insertion into expression vector of choice.

Note: The plasmid DNA was isolated from dam$^+$-methylated *E.coli*. Therefore some restriction sites are blocked by methylation. If you wish to digest the vector using such sites you will need to transform the vector into a dam$^-$ host and make fresh DNA.

The vector backbone contains immediate early promoter of cytomegalovirus ($P_{CMV\ IE}$) for protein expression, SV40 origin for replication in mammalian cells expressing SV40 T-antigen, pUC origin of replication for propagation in *E. coli*, and f1 origin for single-stranded DNA production. SV40 polyadenylation signals (SV40 poly A) direct proper processing of the 3'-end of the reporter mRNA.

SV40 early promoter (P_{SV40}) provides neomycin resistance gene (Neo^r) expression to select stably transfected eukaryotic cells using G418. Bacterial promoter (P) provides kanamycin resistance gene expression (Kan^r) in *E. coli*. Kan^r/Neo^r gene is linked with herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signals.

Expression in mammalian cells

pFusionRed-actinin vector can be transfected into mammalian cells by any known transfection method. CMV promoter provides strong, constitutive expression of the FusionRed-actinin fusion in eukaryotic cells. If required, stable transformants can be selected using G418 [Gorman 1985].

Propagation in *E. coli*

Suitable host strains for propagation in *E. coli* include DH5alpha, HB101, XL1-Blue, and other general purpose strains. Plasmid incompatibility group is pMB1/ColE1. The vector confers resistance to kanamycin (30 μg/ml) to *E. coli* hosts. Copy number in *E. coli* is about 500.

References

Gorman, C. (1985). "High efficiency gene transfer into mammalian cells." In: *DNA cloning: A Practical Approach, Vol. II*. Ed. by Glover. (IRL Press, Oxford, U.K.) Pp. 143–190.

Haas, J. et al. (1996) "Codon usage limitation in the expression of HIV-1 envelope glycoprotein." Curr Biol, 6 (3): 315–324 / pmid: 8805248